

*All cells in the cardiovascular system express one or more subtypes of purine receptors making them possible targets for therapeutic strategies in cardiovascular diseases.*

# Involvement of Purinergic Signaling in Cardiovascular Diseases

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All cells in the heart and blood vessels express one or more subtypes of purine receptors for extracellular adenosine, ATP, ADP, UTP and UDP. P1 receptors, of which there are four subtypes (A<sub>1</sub>, A<sub>2A</sub>, A<sub>2B</sub>, A<sub>3</sub>), are activated by adenosine.<sup>1</sup> There are two distinct families of receptors for purine and pyrimidine nucleotides: P2X ligand-gated ionotropic channels and P2Y metabotropic G-protein-coupled receptors.<sup>2</sup> There are currently eight members of the P2X receptor family (P2X<sub>1-8</sub>) and seven members of the P2Y receptor family (P2Y<sub>1</sub>, P2Y<sub>2</sub>, P2Y<sub>4</sub>, P2Y<sub>6</sub>, P2Y<sub>11</sub>, P2Y<sub>12</sub> and P2Y<sub>13</sub>)<sup>1,3,4</sup>. These subtypes are expressed with some selectivity on different types of cells in the cardiovascular system, and are possible targets for therapeutic strategies in cardiovascular diseases. Figure 1 illustrates the distribution of P1 and P2 receptors in blood vessels.

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## Summary

Purine receptor subtypes are expressed with some selectivity on different types of cells in the cardiovascular system and are possible targets for therapeutic strategies in cardiovascular diseases. The considerable efforts of the clinical biochemists working in this field is leading to a promising emergence of subtype-specific P2 ligands. This will open up new avenues for research into the physiological roles of purine receptors and their therapeutic potential. © 2003 Prous Science. All rights reserved.

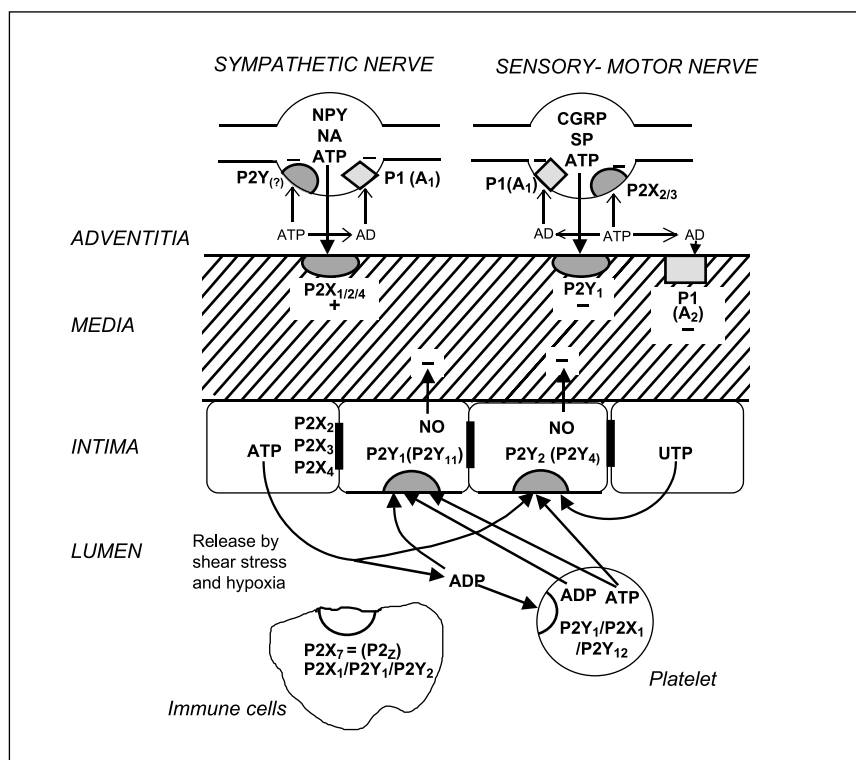
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## Roles of purines and pyrimidines in the cardiovascular system

The extracellular effects of purines in the cardiovascular system have been known for many years.<sup>5-7</sup> The first suggestion that purines acted in a cardioprotective role came with the demonstration that adenosine mediated vasodilatation during hypoxia to increase blood flow and thus maintain oxygen delivery to the heart.<sup>8,9</sup> It is now clear that adenine and pyrimidine nucleotides are released from the heart during hypoxia,<sup>10-12</sup> particularly from endothelial cells,<sup>13,14</sup> and act on P2Y receptors on endothelial cells to produce nitric oxide (NO) and subsequent vasodilatation, before breaking down to adenosine.<sup>15-17</sup> Furthermore, the cardioprotective effects of purine nucleosides and nucleotides involve not only

their actions as potent vasodilators, but also effects on platelet aggregation and trophic actions on smooth muscle and endothelial cells.<sup>18,19</sup> It follows that perturbation of these functions may be involved in some cardiovascular diseases, and indeed there is some evidence for altered levels of expression of P2 receptors and purine receptor signaling in some conditions.

In most blood vessels, P2X<sub>1</sub> receptors are the principal subtype of P2X receptor expressed on the smooth muscle and mediate contraction; they are involved in fast excitatory neurotransmission, being activated by ATP released as a cotransmitter with noradrenaline from perivascular sympathetic nerves.<sup>20</sup> Contractile P2Y<sub>2</sub>, P2Y<sub>4</sub> and P2Y<sub>6</sub> receptors and vasorelaxant A<sub>2</sub> receptors are also expressed on vascular smooth muscle.<sup>1,21</sup> Some vascu-



**Fig. 1.** Short-term (acute) purinergic signaling controlling vascular tone. Schematic illustrating the main receptor subtypes for purine and pyrimidines present in most blood vessels. Perivascular nerves in the adventitia release ATP as cotransmitter: ATP is released with noradrenaline (NA) and neuropeptide Y (NPY) from sympathetic nerves to act on smooth muscle P2X<sub>1</sub>- and in some vessels P2X<sub>2</sub> and P2X<sub>4</sub> purinoceptors, resulting in vasoconstriction; it is released with calcitonin gene-related peptide (CGRP) and substance P (SP) from sensory nerves during 'axon reflex' activity to act on smooth muscle P2Y-purinoceptors resulting in vasodilatation. P1 (A<sub>1</sub>)-purinoceptors on nerve terminals of sympathetic and sensory nerves mediate adenosine (arising from enzymatic breakdown of ATP) modulation of transmitter release. P2X<sub>3</sub> purinoceptors are present on a subpopulation of sensory nerve terminals. P1 (A<sub>2</sub>)-purinoceptors on vascular smooth muscle mediate vasodilatation. Endothelial cells release ATP and UTP during shear stress and hypoxia to act on P2Y<sub>1</sub>, P2Y<sub>2</sub> and sometimes P2Y<sub>4</sub> purinoceptors leading to the production of NO and subsequent vasodilatation. ATP, following its release from aggregating platelets, also acts on these endothelial receptors. Blood-borne platelets possess P2Y<sub>1</sub> and P2Y<sub>12</sub> ADP-selective purinoceptors as well as P2X<sub>1</sub> receptors, while immune cells of various kinds possess P2X<sub>7</sub>, as well as P2X<sub>1</sub>, P2Y<sub>1</sub> and P2X<sub>2</sub> purinoceptors. P2X<sub>2</sub>, P2X<sub>3</sub> and P2X<sub>4</sub> receptors have also recently been identified on endothelial cell membranes. (Reproduced with permission from Burnstock, G. *Development and perspectives of the purinoceptor concept*. *J Auton Pharmacol* 1996, 16: 295-302.)

lar smooth muscle cells, notably those in coronary arteries of a variety of species, also express vasorelaxant P2Y receptors (P2Y<sub>1</sub>-like).<sup>22-26</sup> P2Y and A<sub>2</sub> receptors expressed on smooth muscle cells additionally have roles in control of cell proliferation (Fig. 2).<sup>19</sup>

The vascular endothelium expresses principally P2Y<sub>1</sub>, P2Y<sub>2</sub> and A<sub>2</sub> receptors, which mediate vasorelaxation to ADP, UTP/ATP and adenosine, respectively, following the release of the purines and pyrimidines from the

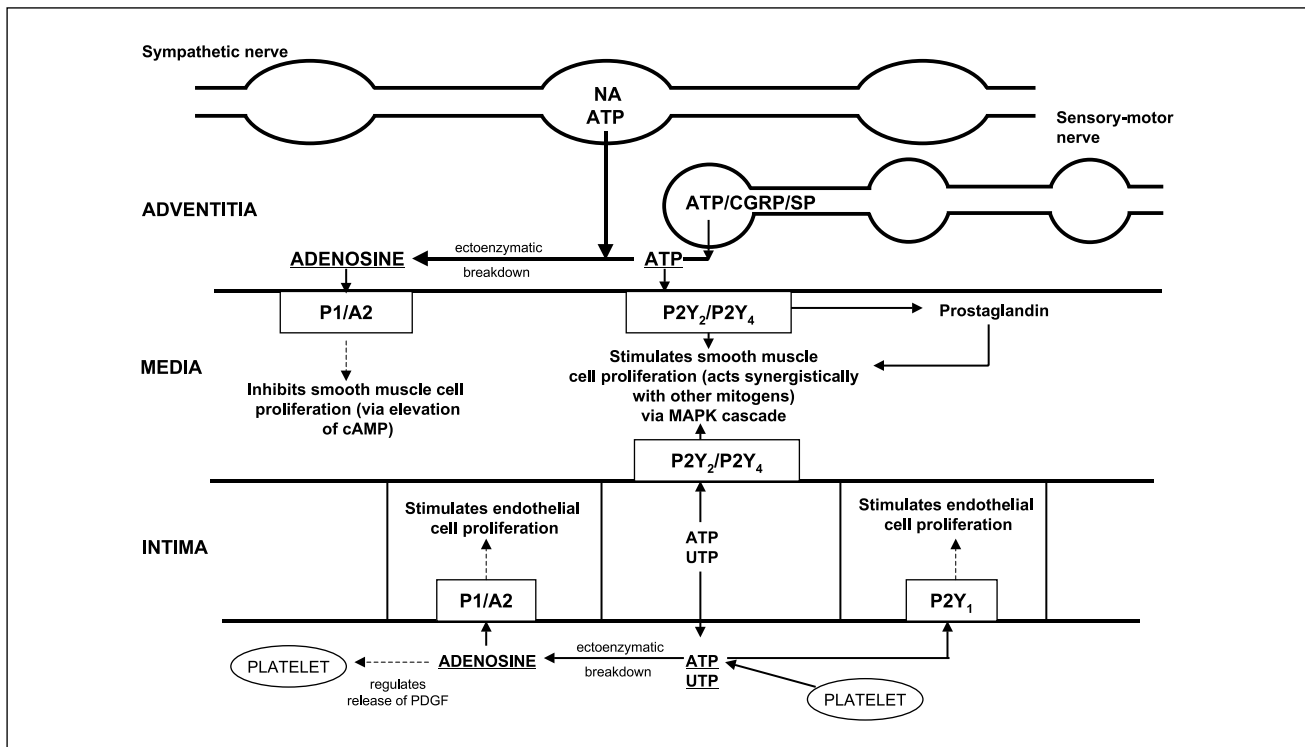
endothelium, platelets and erythrocytes.<sup>1</sup> P2Y and A<sub>2</sub> receptors expressed on endothelial cells are also involved in control of cell proliferation (Fig. 2).<sup>19</sup> Recent evidence suggests that P2X receptors are also expressed on the endothelium.<sup>27-30</sup> There is evidence that shear stress causes Ca<sup>2+</sup> influx via activation of P2X<sub>4</sub> receptors on human endothelial cells.<sup>31-33</sup> The function of these endothelial P2X receptors is uncertain, but roles in cell permeability and adhesion<sup>34</sup> have been proposed.

The nerves in the adventitia of blood vessels express purine receptors. Inhibitory A<sub>1</sub> receptors are expressed on sympathetic and sensory nerves, inhibitory P2Y receptors are expressed on some sympathetic nerves, and heteromeric P2X<sub>2/3</sub> receptors, which may be involved in nociception, have been described on sensory nerves.<sup>1,35</sup>

In the heart, adenosine is a potent coronary vasodilator and causes tachycardia and slows atrioventricular node conductance.<sup>36</sup> It inhibits neutrophil activation, reduces infarct size and preserves endothelial function during inflammation and is also involved in preconditioning via activation of A<sub>1</sub> and A<sub>3</sub> receptors. All four subtypes of adenosine receptor have been localized in the heart;<sup>1,37</sup> the A<sub>1</sub> receptor is associated with the negative cardiac inotropic, chronotropic and dromotropic effects of adenosine, while A<sub>2</sub> receptors mediate coronary vasodilatation.<sup>36,38</sup> Mice lacking the adenosine A<sub>2A</sub> receptor were found to have an increase in heart rate and high blood pressure (as well as being more aggressive and hypoalgesic), identifying a key role of the A<sub>2A</sub> receptor in controlling cardiovascular function.<sup>39</sup>

ATP generally elicits positive inotropic effects, and upon rapid application to cells induces various forms of arrhythmia.<sup>40</sup> It elicits a variety of effects on single cardiomyocytes including activation of nonspecific depolarizing cationic and Cl<sup>-</sup> currents, increases and decreases in Ca<sup>2+</sup> currents, activation of inwardly rectifying currents and outward K<sup>+</sup> currents, an increase in cAMP, and activation of tyrosine kinases to produce Cl<sup>-</sup>/HCO<sub>3</sub><sup>-</sup> exchange and large transient acidosis.<sup>40</sup> Other possible roles of ATP in the heart include hypertrophy, preconditioning and apoptosis. Which of the several subtypes of P2X and P2Y receptors expressed in the heart<sup>41-47</sup> is associated with each of these effects is currently unclear.

The P2Y<sub>12</sub> receptor is found almost exclusively on platelets and is responsible (together with the P2Y<sub>1</sub> and P2X<sub>1</sub>



**Fig. 2.** Schematic of long-term (trophic) actions of purines released from nerves, platelets and endothelial cells (which also release UTP) acting on P2 receptors to stimulate or inhibit cell proliferation. ATP released as a cotransmitter from sympathetic nerves and sensory-motor nerves (during axon reflex activity) stimulates smooth muscle cell proliferation via P2Y<sub>2</sub> and/or P2Y<sub>4</sub> receptors via a MAPK cascade, while adenosine resulting from enzymatic breakdown of ATP acts on P1 (A2) receptors to inhibit cell proliferation (via elevation of cAMP). ATP and UTP released from endothelial cells stimulate both endothelial and smooth muscle cell proliferation via P2Y<sub>1</sub>, P2Y<sub>2</sub> and P2Y<sub>4</sub> receptors. Adenosine resulting from ATP breakdown acts on P1 (A2) receptors to stimulate endothelial cell proliferation and regulate release of PDGF from platelets. (Reproduced with permission from Burnstock, G. *Purinergic signalling and vascular cell proliferation and death*. *Arterioscler Thromb Vasc Biol* 2002, 22: 364-73.)

receptors) for mediating the aggregatory properties of adenine nucleotides.<sup>3,18,48</sup>

### Ectonucleotidases

Once released, ATP is rapidly degraded to ADP, AMP and adenosine by the actions of a family of ectonucleotidases<sup>49-51</sup> and adenosine is further rapidly metabolized via a series of breakdown products to, ultimately, uric acid. The degradation of ATP and adenosine is extremely rapid.<sup>52,53</sup> Ectonucleotidases are situated at the surface of cells and may also be co-released with ATP, for example from sympathetic nerves<sup>54</sup> and endothelial cells.<sup>55</sup> Not only does this limit the actions of ATP at P2 receptors by removing it through enzymatic breakdown, but it can evoke opposite effects via the actions of the breakdown products ADP and adenosine at P2Y and P1 receptors, respectively. When ATP is

released from cells there is likely a cascade of events involving its metabolic breakdown products and activation of multiple subtypes of purine receptors.

### Purinergic signaling in cardiovascular diseases: Therapeutic approaches

Until recently, exploiting cardiovascular purine P2 receptors as therapeutic targets has been limited by the lack of subtype-specific ligands for these receptors. There is evidence, however, that this is changing. For example, P2Y<sub>12</sub> receptor antagonists are an evolving therapeutic strategy as antithrombotics in cardiovascular disease,<sup>56,57</sup> while a highly specific P2Y<sub>1</sub> receptor antagonist, MRS-2279,<sup>58</sup> has recently been developed, holding promise that this or future compounds may find application as antithrombotics, as well as providing new

insights into the physiological role of the P2Y<sub>1</sub> receptor. In addition, TNT-ATP is a powerful antagonist for the P2X<sub>1</sub> receptor that mediates vasoconstriction.<sup>59</sup>

### Antithrombotics

This is a success story in the use of purine receptors as therapeutic targets. ATP and ADP are involved in many of the hemostatic mechanisms that become prominent at sites of vascular injury. When collagen becomes exposed due to endothelial cell damage, platelets begin to adhere to it, aggregate and release ATP; this is broken down to ADP, which promotes further aggregation via activation of P2Y<sub>12</sub> receptors.<sup>3,18,60</sup> ADP can also act on P2Y<sub>1</sub> receptors, either on endothelial cells or on exposed vascular smooth muscle cells, and thereby evoke dilatation. Platelet aggregation is inhibited by ATP and certain

adenine dinucleotides, acting as competitive antagonists on the platelet P2Y<sub>12</sub> receptor, so high local concentrations of these compounds arising as a result of degranulation could serve to control the extent of aggregation. The P2Y<sub>12</sub> receptor is the molecular target of the antithrombotic drugs clopidogrel and ticlopidine<sup>3,56,61-63</sup> and P2Y<sub>1</sub> receptor antagonists are currently being explored as antithrombotic agents by several pharmaceutical companies. A P2X<sub>1</sub> receptor is also present on platelets that modulates calcium influx, but its functional significance is unclear.

A recent study has shown that ATP causes platelet aggregation in whole blood, but not in platelet-rich plasma, implying that ectonucleotidases normally present in whole blood have a significant role in converting the ATP to ADP, which is active at platelet P2Y<sub>1</sub> and P2Y<sub>12</sub> receptors; it is likely that the levels of these ectonucleotidases vary between individuals, with the possibility that levels are low in individuals with impaired blood clotting (S. Heptinstall, personal communication). Increasing or decreasing ATP breakdown could be exploited to alter the balance of purine effects in the cardiovascular system, and in this respect ectonucleotidases are a novel drug target in cardiovascular disease.

### *Supraventricular tachycardia*

Slowing of the heart rate occurs via A<sub>1</sub> receptors on sinoatrial and atrioventricular nodes, causing bradycardia and heart block, respectively, while the inotropic effects include a decrease in atrial contraction and action potential duration.<sup>36</sup> This aspect of A<sub>1</sub> receptor activation has found application in the clinical use of adenosine to treat supraventricular tachycardia,<sup>64</sup> and in the use of adenosine receptor antagonists in the treatment of bradyarrhythmias. ATP has a similar efficacy to adenosine, likely acting via adenosine following metabolic breakdown.<sup>65</sup>

Adenosine has been used as a diagnostic tool of wide QRS complex tachycardias where the mechanism is

uncertain.<sup>66-69</sup> Wide complex ventricular tachycardia is often misdiagnosed as wide complex supraventricular tachycardia with aberrancy, and the use of verapamil in misdiagnosed patients can lead to severe hypotension and cardiac arrest.<sup>67-69</sup> Adenosine can help to differentiate between these conditions because of its specific action on AV nodal conductance.

A recent report has described that the gap junctions in the intercalated disks between human cardiac muscle show some colocalization and some alternate immunostaining for connexin43 and P2X<sub>1</sub> (L. Jiang, personal communication).

### *Ischemia and reperfusion*

During ischemia there is an accumulation of adenosine in the ischemic tissue which may exert protective effects during the reperfusion period via cardiovascular, metabolic and anti-inflammatory effects. Adenosine has, therefore, been used as a cytoprotective agent during myocardial ischemia and reperfusion, although the exact mechanism by which it exerts its beneficial effects is still unclear.<sup>70-72</sup> There is evidence that A<sub>1</sub> receptors are involved, at least in the heart.<sup>73-80</sup> The mechanism may involve A<sub>1</sub> receptor activation of K<sub>ATP</sub> channels, as infarct size reduction after activation of A<sub>1</sub> receptors is abolished by the blockade of K<sub>ATP</sub> channels.<sup>73,81,82</sup>

There is a recent report that in congestive heart failure in rats there is an increase by 2.7-fold and 4.7-fold of P2X<sub>1</sub> and P2Y<sub>2</sub> mRNA levels, respectively.<sup>83</sup>

### *Hypertension, diabetes and blood pressure regulation*

There is evidence that ATP plays a significantly greater role as a sympathetic cotransmitter in spontaneously hypertensive rats.<sup>84-86</sup> Defective prejunctional P1 receptor-mediated modulation has been claimed to contribute to enhanced sympathetic neurotransmission in hypertension.<sup>87-89</sup> Constriction of the isolated kidney to renal nerve stimulation was increased in

spontaneously hypertensive rats compared with controls and appeared to be entirely due to ATP released from sympathetic nerves.<sup>90</sup> In the kidney of a transgenic, hypertensive model, the glomeruli show an abundance of P2X<sub>7</sub> immunostaining and a similar expression of P2X<sub>7</sub> receptors has been found in the glomeruli of diabetic rats with kidney damage.<sup>91</sup>

Intravenous infusion of ATP in humans has been investigated with regard to its therapeutic use in patients with chronic obstructive pulmonary disease.<sup>92</sup> Pulmonary hypertension can be a major problem after thoracic surgery in patients with chronic obstructive pulmonary disease. Low-dose intravenous ATP or adenosine have been used as vasodilators under these conditions as they have been shown to have predominantly pulmonary vasodilating effects,<sup>93,94</sup> an advantage over vasodilators with predominant systemic effects. Pulmonary hypertension can also be a serious problem in children after surgical repair of congenital heart defects, and in this respect intravenous ATP produced a reduction or disappearance of the pulmonary hypertensive crisis without a change in mean systemic arterial pressure and systemic vascular resistance.<sup>95</sup>

Because it is a potent vasodilator in most vascular beds, ATP has been investigated with respect to pharmacologically induced hypotension, which is frequently used during surgery to optimize operating conditions and decrease chances of hemorrhage.<sup>69,96</sup> The advantages of ATP-induced hypotension are the absence of tachyphylaxis, the stability of blood pressure with well-preserved hemodynamics, fast onset and rapid reversibility with no rebound hypotension.

### *Atherosclerosis and restenosis*

Adenosine and ATP have a number of cardiovascular protective effects in addition to vasodilatation, including the promotion of endothelial and smooth muscle cell proliferation and an increase in the expression of vascu-

lar endothelial growth factor (VEGF) mRNA (Fig. 2), which play an important role in the development of intimal thickening during arterial diseases, such as atherosclerosis, and in restenosis after angioplasty, and in the growth of new vessels that takes place during wound healing and in tumors.<sup>19</sup> Hypoxia is an important stimulus to vascular growth and it is believed that adenosine and ATP, both of which are released from cells during hypoxia, have important roles as mediators of blood vessel growth.

Endothelial cell proliferation is mediated by A<sub>2A</sub> and A<sub>2B</sub> receptors, and some of the mitogenic effects are mediated via the modulation of VEGF signalling. There is evidence that A<sub>2B</sub> receptors inhibit the growth of human aortic smooth muscle cells.<sup>97</sup> ATP and ADP stimulate DNA synthesis and cell proliferation of porcine aortic smooth muscle cells<sup>98</sup> via activation of P<sub>2Y</sub> receptors. UTP also has powerful mitogenic actions on vascular smooth muscle, and since the mitogenic effects of UTP and ATP were approximately equipotent,<sup>99,100</sup> this would suggest that the receptor involved is either a P<sub>2Y</sub><sub>2</sub> or P<sub>2Y</sub><sub>4</sub> subtype. There was upregulation of vascular smooth muscle P<sub>2Y</sub><sub>2</sub> receptors by MAPK kinase-dependent growth factor, which the authors suggested may be important in atherosclerosis and neointimal formation after balloon angioplasty.<sup>47</sup> ATP and ADP have also been shown to stimulate endothelial cell migration and proliferation,<sup>101,102</sup> probably via P<sub>2Y</sub> receptors.

Apoptotic cell death is recognized to occur in a number of vascular diseases, including atherosclerosis, restenosis and hypertension.<sup>103,104</sup> ATP releases histamine from mast cells and releases inflammatory cytokines such as IL-1 from immune cells.<sup>105</sup> In addition, occupation of P<sub>2</sub> receptors leads to prostaglandin and COX-2 synthesis,<sup>106</sup> both involved in inflammatory processes. Vascular endothelial cells are continuously exposed to variations in blood flow, which plays an important role in vessel growth or regression

and in the local development of atherosclerosis. The shear stress leads to a substantial release of ATP and UTP from endothelial cells<sup>16</sup> and these purines might mediate alterations in the balance between proliferation and apoptosis.<sup>107</sup>

In restenosis following balloon angioplasty, there is a peak in the proliferation and apoptosis of vascular smooth muscle at about 14 days.<sup>108</sup> The first balloon inflation during coronary angioplasty is a preconditioning stimulus leading to a decrease in ischemia in later inflations; intracoronary adenosine administration before coronary angioplasty modifies the preconditioning effect of the first inflation.<sup>109</sup>

### Vascular inflammation

Adenosine is released at sites of inflammation and has antiinflammatory effects via multiple mechanisms. It inhibits neutrophil rolling and adhesion to vascular endothelium, decreases oxygen-free radical production by neutrophils via activation of A<sub>2A</sub> receptors and also has effects on endothelial cell permeability via A<sub>1</sub> and A<sub>2A</sub> receptors.<sup>110</sup> Adenosine also inhibits macrophage production of the proinflammatory cytokine tumor necrosis factor- $\alpha$  (TNF $\alpha$ ) and suppresses TNF $\alpha$  mRNA expression and plasma levels.<sup>110</sup> A recent study in knockout A<sub>2A</sub> mice showed an important role of A<sub>2A</sub> receptors in preventing liver damage in inflammation and maintaining lower levels of serum TNF $\alpha$ .<sup>111</sup> P<sub>2</sub> receptors can also be involved in inflammation, as ATP induces a localized inflammatory response in rat paw.<sup>112</sup>

### Vascular pain

It has been suggested that in vascular pain, including migraine, angina, pelvic pain and ischemic pain, that ATP released from endothelial cells during the reactive hyperemia following vasospasm (not associated with pain) diffuses through the wall of microvessels to reach P<sub>2X</sub><sub>3</sub> receptors on sensory perivascular nerves to initiate impulses that travel via the spinal cord to pain centers in the brain.<sup>35</sup> Furthermore, ATP activation of P<sub>2Y</sub><sub>1</sub>

receptors potentiates vanilloid VR1 currents evoked by capsaicin or protons and reduces the temperature threshold for activation of VR1.<sup>113,114</sup>

Adenosine alleviates neuropathic pain, hyperalgesia and allodynia,<sup>115,116</sup> an action which may be mediated by inhibitory A<sub>1</sub> receptors expressed on sensory nerves. Intravenous ATP in mice was found to have analgesic activity on hot plate and phenyl quinone-induced stretching assays.<sup>117-119</sup>

## Conclusions

There is evidence that purines contribute to a number of processes involved in normal cardiovascular function, and that disturbances in purinergic signaling are involved in some cardiovascular diseases. As all cells in the cardiovascular system express one or more types of purine receptor, this raises the possibility that purine receptors may be potential targets in cardiovascular disease. Indeed, antagonists of P<sub>2Y</sub><sub>12</sub> receptors are now used as antithrombotics, and P<sub>1</sub> agonists for supraventricular tachycardia in humans. It is an advantage that there is some selectivity in the subtypes of P<sub>1</sub> and P<sub>2</sub> receptors that are expressed on different structures in the heart and blood vessels. However, the development and use of ligands with good subtype specificities that do not degrade *in vivo* is needed to exploit this. The considerable efforts of the clinical biochemists working in this field is leading to a promising emergence of subtype-specific P<sub>2</sub> ligands. This will open up new avenues for research into the physiological roles of purine receptors and their therapeutic potential.

## References

1. Ralevic, V. and Burnstock, G. *Receptors for purines and pyrimidines*. Pharmacol Rev 1998, 50: 413–92.
2. Abbracchio, M.P. and Burnstock, G. *Purinoceptors: Are there families of P<sub>2X</sub> and P<sub>2Y</sub> purinoceptors?* Pharmacol Ther 1994, 64: 445–75.
3. Hollopeter, G., Jantzen, H.-M., Vincent, D. et al. *Identification of the platelet ADP receptor targeted by antithrombotic drugs*. Nature 2001, 409: 202–7.

4. Zhang, F.L., Luo, L., Gustafson, E. et al. *P2Y<sub>13</sub>: Identification and characterization of a novel G $\alpha$ phai-coupled ADP receptor from human and mouse*. J Pharmacol Exp Ther 2002, 301: 705–13.
5. Drury, A.N. *The physiological activity of nucleic acid and its derivatives*. Physiol Rev 1936, 16: 292–325.
6. Gillespie, J.H. *The biological significance of the linkages in adenosine triphosphoric acid*. J Physiol (Lond) 1934, 80: 345–59.
7. Green, H.N. and Stoner, H.B. *Biological Actions of the Adenine Nucleotides*. Lewis, London 1950.
8. Berne, R.M. *Cardiac nucleotides in hypoxia: Possible role in regulation of coronary blood flow*. Am J Physiol 1963, 204: 317–22.
9. Gerlach, E., Deuticke, B. and Dreisbach, R.H. *Der-nucleotid-abbau in herzmuskel bei sauerstoffmangel und seine mögliche bedeutung für die coronardurchblutung*. Naturwissenschaften 1963, 50: 228–9.
10. Paddle, B.M. and Burnstock, G. *Release of ATP from perfused heart during coronary vasodilatation*. Blood Vessels 1974, 11: 110–9.
11. Forrester, T. and Williams, C.A. *Release of adenosine triphosphate from isolated adult heart cells in response to hypoxia*. J Physiol 1977, 268: 371–90.
12. Clemens, M.G. and Forrester, T. *Appearance of adenosine triphosphate in the coronary sinus effluent from isolated working rat heart in response to hypoxia*. J Physiol 1981, 312: 143–58.
13. Bodin, P. and Burnstock, G. *Synergistic effect of acute hypoxia on flow-induced release of ATP from cultured endothelial cells*. Experientia 1995, 51: 256–9.
14. Bodin, P., Milner, P., Winter, R. and Burnstock, G. *Chronic hypoxia changes the ratio of endothelin to ATP release from rat aortic endothelial cells exposed to high flow*. Proc R Soc Lond B Biol Sci 1992, 247: 131–5.
15. Burnstock, G. *Hypoxia, endothelium and purines*. Drug Dev Res 1993, 28: 301–5.
16. Burnstock, G. *Release of vasoactive substances from endothelial cells by shear stress and purinergic mechanosensory transduction*. J Anat 1999, 194: 335–42.
17. Saïag, B., Bodin, P., Shacoori, V., Catheline, M., Rault, B. and Burnstock, G. *Uptake and flow-induced release of uridine nucleotides from isolated vascular endothelial cells*. Endothelium 1995, 2: 279–85.
18. Boarder, M.R. and Hourani, S.M.O. *The regulation of vascular function by P2 receptors: multiple sites and multiple receptors*. Trends Pharmacol Sci 1998, 19: 99–107.
19. Burnstock, G. *Purinergic signalling and vascular cell proliferation and death*. Arterioscl Thromb Vasc Biol 2002, 22: 364–73.
20. Burnstock, G. *Co-transmission. The fifth Heymans memorial lecture - Ghent, February 17, 1990*. Arch Int Pharmacodyn Ther 1990, 304: 7–33.
21. Bodin, P. and Burnstock, G. *Evidence that release of ATP from endothelial cells during increased shear stress is vesicular*. J Cardiovasc Pharmacol 2001, 38: 900–8.
22. Keef, K.D., Pasco, J.S. and Eckman, D.M. *Purinergic relaxation and hyperpolarization in guinea pig and rabbit coronary artery: Role of the endothelium*. J Pharmacol Exp Ther 1992, 260: 592–600.
23. Corr, L. and Burnstock, G. *Analysis of P<sub>2</sub>-purinoceptor subtypes on the smooth muscle and endothelium of rabbit coronary artery*. J Cardiovasc Pharmacol 1994, 23: 709–15.
24. Simonsen, U., Garcia-Sacristan, A. and Prieto, D. *Involvement of ATP in the non-adrenergic non-cholinergic inhibitory neurotransmission of lamb isolated coronary small arteries*. Br J Pharmacol 1997, 120: 411–20.
25. Ströbaek, D., Olesen, S.-P., Christoffersen, P. and Dissing, S. *P<sub>2</sub>-purinoceptor-mediated formation of inositol phosphates and intracellular Ca<sup>2+</sup> transients in human coronary artery smooth muscle cells*. Br J Pharmacol 1995, 118: 1645–52.
26. Alexander, S.P., Hawkes, J., Patel, S. and Ralevic, V. *Characterisation of the novel P<sub>2</sub> receptor antagonist MRS2179 in the porcine isolated coronary artery*. Brit J Pharmacol Abstr Submitted 2002.
27. Hansen, M.A., Dutton, J.L., Balcar, V.J., Barden, J.A. and Bennett, M.R. *P<sub>2X</sub> (purinergic) receptor distributions in rat blood vessels*. J Auton Nerv Syst 1999, 75: 147–55.
28. Loesch, A. and Burnstock, G. *Ultrastructural localisation of ATP-gated P<sub>2X</sub> receptor immunoreactivity in vascular endothelial cells in rat brain*. Endothelium 2000, 7: 93–8.
29. Glass, R. and Burnstock, G. *Immunohistochemical identification of cells expressing ATP-gated cation channels (P<sub>2X</sub> receptors) in the adult rat thyroid*. J Anat 2001, 198: 569–79.
30. Schwiebert, L.M., Rice, W.C., Kudlow, B.A., Taylor, A.L. and Schwiebert, E.M. *Extracellular ATP signaling and P<sub>2X</sub> nucleotide receptors in monolayers of primary human vascular endothelial cells*. Am J Physiol Cell Physiol 2002, 282: C289–301.
31. Yamamoto, K., Korenaga, R., Kamiya, A. and Ando, J. *Fluid shear stress activates Ca<sup>2+</sup> influx into human endothelial cells via P<sub>2X</sub> purinoceptors*. Circ Res 2000, 87: 385–91.
32. Korenaga, R., Yamamoto, K., Ohura, N., Sokabe, T., Kamiya, A. and Ando, J. *Sp1-mediated downregulation of P<sub>2X</sub> receptor gene transcription in endothelial cells exposed to shear stress*. Am J Physiol Heart Circ Physiol 2001, 280: H2214–H21.
33. Yamamoto, K., Korenaga, R., Kamiya, A., Qi, Z., Sokabe, M. and Ando, J. *P<sub>2X</sub> receptors mediate ATP-induced calcium influx in human vascular endothelial cells*. Am J Physiol Heart Circ Physiol 2000, 279: H285–H292.
34. Glass, R., Loesch, A., Bodin, P. and Burnstock, G. *P<sub>2X</sub> and P<sub>2X</sub> receptors associate with VE-cadherin in human endothelial cells*. Cell Mol Life Sci 2002, 59: 870–81.
35. Burnstock, G. *A unifying purinergic hypothesis for the initiation of pain*. Lancet 1996, 347: 1604–5.
36. Olsson, R.A. and Pearson, J.D. *Cardiovascular purinoceptors*. Physiol Rev 1990, 70: 761–845.
37. Dixon, A.K., Gubitz, A.K., Sirinathsinghji, D.J.S., Richardson, P.J. and Freeman, T.C. *Tissue distribution of adenosine receptor mRNAs in the rat*. Br J Pharmacol 1996, 118: 1461–8.
38. Collis, M.G. and Hourani, S.M. *Adenosine receptor subtypes*. Trends Pharmacol Sci 1993, 14: 360–6.
39. Ledent, C., Vaugeois, J.M., Schiffmann, S.N. et al. *Aggressiveness, hypoalgesia and high blood pressure in mice lacking the adenosine A<sub>2a</sub> receptor*. Nature 1997, 388: 674–8.
40. Vassort, G. *Adenosine 5'-triphosphate: A P<sub>2</sub>-purinergic agonist in the myocardium*. Physiol Rev 2001, 81: 767–806.
41. Webb, T.E., Boluyt, M.O. and Barnard, E.A. *Molecular biology of P<sub>2Y</sub> purinoceptors: Expression in rat heart*. J Auton Pharmacol 1996, 16: 303–7.
42. Garcia-Guzman, M., Soto, F., Laube, B. and Stühmer, W. *Molecular cloning and functional expression of a novel rat heart P<sub>2X</sub> purinoceptor*. FEBS Lett 1996, 388: 123–7.
43. Garcia-Guzman, M., Soto, F., Gomez-Hernandez, J.M., Lund, P.E. and Stühmer, W. *Characterization of recombinant human P<sub>2X</sub> receptor reveals pharmacological differences to the rat homologue*. Mol Pharmacol 1997, 51: 109–18.
44. Soto, F., Garcia, G.M., Gomez-Hernandez, J.M., Hollmann, M., Karschin, C. and Stühmer, W. *P<sub>2X</sub>: An ATP-activated ionotropic receptor cloned from rat brain*. Proc Natl Acad Sci USA 1996, 93: 3684–8.
45. Bogdanov, Y., Rubino, A., Burnstock, G. *Characterisation of subtypes of the P<sub>2X</sub> and P<sub>2Y</sub> families of receptors in the foetal human heart*. Life Sci 1998, 62: 697–703.
46. Nori, S., Fumagalli, L., Bo, X., Bogdanov, Y. and Burnstock, G. *Coexpression of mRNAs for P<sub>2X</sub>, P<sub>2X</sub> and P<sub>2X</sub> receptors in rat vascular smooth muscle: An in situ hybridization and RT-PCR study*. J Vasc Res 1998, 35: 179–85.
47. Hou, M., Moller, S., Edvinsson, L. and Erlinge, D. *MAPKK-dependent growth factor-induced upregulation of P<sub>2Y</sub> receptors in vascular smooth muscle cells*. Biochem Biophys Res Commun 1999, 258: 648–52.

48. Di Virgilio, F. and Solini, A. *P2 receptors: New potential players in atherosclerosis*. *Br J Pharmacol* 2002, 135: 831–42.
49. Zimmermann, H. *5'-Nucleotidase: Molecular structure and functional aspects*. *Biochem J* 1992, 285: 345–65.
50. Ziganshin, A.U., Hoyle, C.H.V. and Burnstock, G. *Ecto-enzymes and metabolism of extracellular ATP*. *Drug Dev Res* 1994, 32: 134–46.
51. Zimmermann, H. and Braun, N. *Extracellular metabolism of nucleotides in the nervous system*. *J Auton Pharmacol* 1996, 16: 397–400.
52. Ryan, J.W. and Smith, U. *Metabolism of adenosine 5'-monophosphate during circulation through the lungs*. *Trans Assoc Am Physicians* 1971, 84: 297–306.
53. Moser, G.H., Schrader, J. and Deussen, A. *Turnover of adenosine in plasma of human and dog blood*. *Am J Physiol* 1989, 256: C799–C806.
54. Todorov, L.D., Mihaylova Todorova, S., Westfall, T.D. et al. *Neuronal release of soluble nucleotidases and their role in neurotransmitter inactivation*. *Nature* 1997, 387: 76–9.
55. Yegutkin, G., Bodin, P. and Burnstock, G. *Effect of shear stress on the release of soluble ecto-enzymes ATPase and 5'-nucleotidase along with endogenous ATP from vascular endothelial cells*. *Br J Pharmacol* 2000, 129: 921–6.
56. Storey, F. *The P2Y12 receptor as a therapeutic target in cardiovascular disease*. *Platelets* 2001, 12: 197–209.
57. Yusuf, S., Zhao, F., Mehta, S.R. et al. *The clopidogrel in unstable angina to prevent recurrent events trial investigators: Effects of clopidogrel in addition to aspirin in patients with acute coronary syndromes without ST-segment elevation*. *N Engl J Med* 2001, 345: 494–502.
58. Boyer, J.L., Adams, M., Ravi, R.G., Jacobson, K.A. and Harden, T.K. *2-Chloro-N(6)-methyl-(N)-methanocarba-2'-deoxyadenosine-3',5'-bisphosphate is a selective high affinity P2Y(1) receptor antagonist*. *Br J Pharmacol* 2002, 135: 2004–10.
59. Virginio, C., Robertson, G., Surprenant, A. and North, R.A. *Trinitrophenyl-substituted nucleotides are potent antagonists selective for P2X<sub>1</sub>, P2X<sub>3</sub>, and heteromeric P2X<sub>2/3</sub> receptors*. *Mol Pharmacol* 1998, 53: 969–73.
60. Gachet, C. *ADP receptors of platelets and their inhibition*. *Thromb Haemost* 2001, 86: 222–32.
61. Boyer, J.L., Lazarowski, E.R., Chen, X.H. and Harden, T.K. *Identification of a P2Y-purinergic receptor that inhibits adenyl cyclase*. *J Pharmacol Exp Ther* 1993, 267: 1140–6.
62. Daniel, J.L., Dangelmaier, C., Jin, J., Ashby, B., Smith, J.B. and Kunapuli, S.P. *Molecular basis for ADP-induced platelet activation*. *J Biol Chem* 1998, 273: 2024–9.
63. Foster, C.J., Prosser, D.M., Agans, J.M. et al. *Molecular identification and characterization of the platelet ADP receptor targeted by thienopyridine antithrombotic drugs*. *J Clin Invest* 2001, 107: 1591–8.
64. Rankin, A.C., Brooks, R., Ruskin, J.N. and McGovern, B.A. *Adenosine and the treatment of supraventricular tachycardia*. *Am J Med* 1992, 92: 655–64.
65. Rankin, A.C., Oldroyd, K.G., Chong, E., Dow, J.W., Rae, A.P. and Cobbe, S.M. *Adenosine or adenosine triphosphate for supraventricular tachycardias? Comparative double-blind randomized study in patients with spontaneous or inducible arrhythmias*. *Am Heart J* 1990, 119: 316–23.
66. Griffith, M.J., Linker, N.J., Ward, D.E. and Camm, A.J. *Adenosine in the diagnosis of broad complex tachycardia*. *Lancet* 1988, 1: 672–5.
67. Rankin, A.C., Oldroyd, K.G., Chong, E., Rae, A.P. and Cobbe, S.M. *Value and limitations of adenosine in the diagnosis and treatment of narrow and broad complex tachycardias*. *Br Heart J* 1989, 62: 195–203.
68. Sharma, A.D., Klein, G.J. and Yee, R. *Intra-venous adenosine triphosphate during wide QRS complex tachycardia: Safety, therapeutic efficacy, and diagnostic utility*. *Am J Med* 1990, 88: 337–43.
69. Agteresch, H.J., Dagnelie, P.C., van den Berg, J.W.O. and Wilson, J.H. *Adenosine triphosphate: Established and potential clinical applications*. *Drugs* 1999, 58: 211–32.
70. Leeser, M.A., Stoddard, M., Ahmed, M., Broadbent, J. and Bolli, R. *Preconditioning of human myocardium with adenosine during coronary angioplasty*. *Circulation* 1997, 95: 2500–7.
71. Liang, B.T. and Jacobson, K.A. *Adenosine and ischemic preconditioning*. *Curr Pharm Des* 1999, 5: 1029–41.
72. Sommerschild, H.T. and Kirkeboen, K.A. *Adenosine and cardioprotection during ischaemia and reperfusion—an overview*. *Acta Anaesthesiol Scand* 2000, 44: 1038–55.
73. Grover, G.J., Sleph, P.G. and Dzwonczyk, S. *Role of myocardial ATP-sensitive potassium channels in mediating preconditioning in the dog heart and their possible interaction with adenosine A1-receptors*. *Circulation* 1992, 86: 1310–6.
74. Tsuchida, A., Liu, G.S., Wilborn, W.H. and Downey, J.M. *Pretreatment with the adenosine A1 selective agonist, 2-chloro-N6-cyclopentyladenosine (CCPA), causes a sustained limitation of infarct size in rabbits*. *Cardiovasc Res* 1993, 27: 652–6.
75. Tsuchida, A., Thompson, R., Olsson, R.A. and Downey, J.M. *The anti-infarct effect of an adenosine A1-selective agonist is diminished after prolonged infusion as is the cardioprotective effect of ischaemic preconditioning in rabbit heart*. *J Mol Cell Cardiol* 1994, 26: 303–11.
76. Yao, Z. and Gross, G.J. *Glibenclamide antagonizes adenosine A1 receptor-mediated cardioprotection in stunned canine myocardium*. *Circulation* 1993, 88: 235–44.
77. Lee, Y.M., Sheu, J.R. and Yen, M.H. *BN-063, a newly synthesized adenosine A1 receptor agonist, attenuates myocardial reperfusion injury in rats*. *Eur J Pharmacol* 1995, 279: 251–6.
78. Lasley, R.D. and Mentzer, R.M. Jr. *Protective effects of adenosine in the reversibly injured heart*. *Ann Thorac Surg* 1995, 60: 843–6.
79. Strickler, J., Jacobson, K.A. and Liang, B.T. *Direct preconditioning of cultured chick ventricular myocytes. Novel functions of cardiac adenosine A2a and A3 receptors*. *J Clin Invest* 1996, 98: 1773–9.
80. Matherne, G.P., Linden, J., Byford, A.M., Gauthier, N.S. and Headrick, J.P. *Transgenic A1 adenosine receptor overexpression increases myocardial resistance to ischemia*. *Proc Natl Acad Sci USA* 1997, 94: 6541–6.
81. Van Winkle, D.M., Chien, G.L., Wolff, R.A., Soifer, B.E., Kuzume, K. and Davis, R.F. *Cardioprotection provided by adenosine receptor activation is abolished by blockade of the K<sub>ATP</sub> channel*. *Am J Physiol* 1994, 266: H829–39.
82. Mizumura, T., Auchampach, J.A., Linden, J., Bruns, R.F. and Gross, G.J. *PD 81,723, an allosteric enhancer of the A1 adenosine receptor, lowers the threshold for ischemic preconditioning in dogs*. *Circ Res* 1996, 79: 415–23.
83. Hou, M., Malmisjo, M., Moller, S. et al. *Increase in cardiac P2X<sub>1</sub>- and P2Y<sub>2</sub>-receptor mRNA levels in congestive heart failure*. *Life Sci* 1999, 65: 1195–206.
84. Vidal, M., Hicks, P.E. and Langer, S.Z. *Differential effects of a,b-methylene ATP on responses to nerve stimulation in SHR and WKY tail arteries*. *Naunyn Schmiedeberg Arch Pharmacol* 1986, 332: 384–90.
85. Bulloch, J.M. and McGrath, J.C. *Evidence for increased purinergic contribution in hypertensive blood vessels exhibiting co-transmission*. *Br J Pharmacol* 1992, 107: 145P.
86. Brock, J.A. and Van Helden, D.F. *Enhanced excitatory junction potentials in mesenteric arteries from spontaneously hypertensive rats*. *Pflugers Arch* 1995, 430: 901–8.
87. Kamikawa, Y., Cline, W.H. and Su, C. *Diminished purinergic modulation of the vascular adrenergic neurotransmission in spontaneously hypertensive rats*. *Eur J Pharmacol* 1980, 66: 347–53.

88. Kubo, T., Su, C. *Effects of adenosine on [<sup>3</sup>H]norepinephrine release from perfused mesenteric arteries of SHR and renal hypertensive rats*. Eur J Pharmacol 1983, 87: 349–52.
89. Illes, P., Rickmann, H., Brod, I., Bucher, B. and Stoclet, J.-C. *Subsensitivity of presynaptic adenosine A<sub>1</sub>-receptors in caudal arteries of spontaneously hypertensive rats*. Eur J Pharmacol 1989, 174: 237–51.
90. Rump, L.C., Wilde, K. and Schollmeyer, P. *Prostaglandin E<sub>2</sub> inhibits noradrenaline release and purinergic pressor responses to renal nerve stimulation at 1 Hz in isolated kidneys of young spontaneously hypertensive rats*. J Hypertens 1990, 8: 897–908.
91. Chan, C.M., Hillman, K., Moss, S., Unwin, R.J. and Burnstock, G. *Putative cytolytic P<sub>2</sub>X<sub>7</sub> receptor in diabetic rat kidney and its stable transfection in lymphocytes*. J Amer Soc Nephrol 1998, 9[Prog. & Abstr.]: 420A.
92. Gaba, S.J.M., Bourgouin-Karaoui, D., Dujols, P., Michel, F.B. and Prefaut, C. *Effects of adenosine triphosphate on pulmonary circulation in chronic obstructive pulmonary disease. ATP: A pulmonary vasoregulator?* Am Rev Respir Dis 1986, 134: 1140–4.
93. Gaba, S.J. and Prefaut, C. *Comparison of pulmonary and systemic effects of adenosine triphosphate in chronic obstructive pulmonary disease-ATP: A pulmonary controlled vasoregulator?* Eur Respir J 1990, 3: 450–5.
94. Fullerton, D.A., Jagers, J., Jones, S.D., Brown, J.M. and McIntyre, R.C. Jr. *Adenosine for refractory pulmonary hypertension*. Ann Thorac Surg 1996, 62: 874–7.
95. Brook, M.M., Fineman, J.R., Bolinger, A.M., Wong, A.F., Heymann, M.A. and Soifer, S.J. *Use of ATP-MgCl<sub>2</sub> in the evaluation and treatment of children with pulmonary hypertension secondary to congenital heart defects*. Circ 1994, 90: 1287–93.
96. Aso, Y., Tajima, A., Suzuki, K. et al. *Intraoperative blood pressure control by ATP in pheochromocytoma*. Urology 1986, 27: 512–20.
97. Dubey, R.K., Gillespie, D.G. and Jackson, E.K. *Adenosine inhibits collagen and total protein synthesis in vascular smooth muscle cells*. Hypertension 1999, 33: 190–4.
98. Wang, D.J., Huang, N.N., Heppel, L.A. *Extracellular ATP and ADP stimulate proliferation of porcine aortic smooth muscle cells*. J Cell Physiol 1992, 153: 221–33.
99. Erlinge, D., You, J., Wahlestedt, C. and Edvinsson, L. *Characterisation of an ATP receptor mediating mitogenesis in vascular smooth muscle cells*. Eur J Pharmacol 1995, 289: 135–49.
100. Malam-Souley, R., Seye, C., Gadeau, A.P. et al. *Nucleotide receptor P<sub>2u</sub> partially mediates ATP-induced cell cycle progression of aortic smooth muscle cells*. J Cell Physiol 1996, 166: 57–65.
101. McAuslan, B.R., Reilly, W.G., Hannan, G.N. and Gole, G.A. *Angiogenic factors and their assay: Activity of formyl methionyl leucyl phenylalanine, adenosine diphosphate, heparin, copper, and bovine endothelium stimulating factor*. Microvasc Res 1983, 26: 323–38.
102. Van Coevorden, A., Roger, P. and Boeynaems, J.-M. *Mitogenic action of adenosine nucleotides and nucleosides on aortic endothelial cells*. Thromb Haemost 1989, 62: 190.
103. Thomas, W.A., Reiner, J.M., Florentin, F.A., Lee, K.T. and Lee, W.M. *Population dynamics of arterial smooth muscle cells. V. Cell proliferation and cell death during initial 3 months in atherosclerotic lesions induced in swine by hypercholesterolemic diet and intimal trauma*. Exp Mol Pathol 1976, 24: 360–74.
104. Mallat, Z. and Tedgui, A. *Apoptosis in the vasculature: Mechanisms and functional importance*. Br J Pharmacol 2000, 130: 947–62.
105. Di Virgilio, F., Falzoni, S., Mutini, C., Sanz, J.M. and Chiozzi, P. *Purinergic P<sub>2</sub>X<sub>7</sub> receptor: A pivotal role in inflammation and immunomodulation*. Drug Dev Res 1998, 45: 207–13.
106. Brambilla, R., Burnstock, G., Bonazzi, A., Ceruti, S., Cattabeni, F. and Abbracchio, M.P. *Cyclooxygenase-2 mediates P<sub>2</sub>Y receptor-induced reactive astrogliosis. Special Report*. Br J Pharmacol 1999, 126: 563–7.
107. Kaiser, D., Freyberg, M.A. and Friedl, P. *Lack of hemodynamic forces triggers apoptosis in vascular endothelial cells*. Biochem Biophys Res Commun 1997, 231: 586–90.
108. Han, D.K., Haudenschild, C.C., Hong, M.K., Tinkle, B.T., Leon, M.B. and Liau, G. *Evidence for apoptosis in human atherosclerosis and in a rat vascular injury model*. Am J Pathol 1995, 147: 267–77.
109. Kerensky, R.A., Kutcher, M.A., Braden, G.A., Applegate, R.J., Solis, G.A. and Little, W.C. *The effects of intracoronary adenosine on preconditioning during coronary angioplasty*. Clin Cardiol 1995, 18: 91–6.
110. Williams, M. and Jarvis, M.F. *Purinergic and pyrimidineric receptors as potential drug targets*. Biochem Pharmacol 2000, 59: 1173–85.
111. Ohta, A. and Sitkovsky, M. *Role of G-protein-coupled adenosine receptors in down-regulation of inflammation and protection from tissue damage*. Nature 2001, 414: 916–20.
112. Ziganshina, L.E., Ziganshin, A.U., Hoyle, C.H.V. and Burnstock, G. *Acute paw oedema formation induced by ATP: Re-evaluation of the mechanisms involved*. Inflamm Res 1996, 45: 96–102.
113. Tominaga, M., Wada, M. and Masu, M. *Potentiation of capsaicin receptor activity by metabotropic ATP receptors as a possible mechanism for ATP-evoked pain and hyperalgesia*. Proc Natl Acad Sci USA 2001, 98: 6951–6.
114. Numazaki, M., Tominaga, T., Toyooka, H. and Tominaga, M. *Direct phosphorylation of capsaicin receptor VR1 by protein kinase Cε and identification of two target serine residues*. J Biol Chem 2002, 277: 13375–8.
115. Belfrage, M., Sollevi, A., Segerdahl, M., Sjolund, K.F. and Hansson, P. *Systemic adenosine infusion alleviates spontaneous and stimulus evoked pain in patients with peripheral neuropathic pain*. Anesth Analg 1995, 81: 713–7.
116. Sollevi, A., Belfrage, M., Lundeberg, T., Segerdahl, M. and Hansson, P. *Systemic adenosine infusion: A new treatment modality to alleviate neuropathic pain*. Pain 1995, 61: 155–8.
117. Sawynok, J. and Sweeney, M.I. *The role of purines in nociception*. Neurosci 1989, 32: 557–69.
118. Sawynok, J., Sweeney, M.I. and White, T.D. *Adenosine release may mediate spinal analgesia by morphine*. Trends Pharmacol Sci 1989, 10: 186–9.
119. Gomaa, A.A. *Characteristics of analgesia induced by adenosine triphosphate*. Pharmacol Toxicol 1987, 61: 199–202.
120. Burnstock, G. *Development and perspectives of the purinoceptor concept*. J Auton Pharmacol 1996, 16: 295–302.

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