

Primers, PCR conditions, purification of PCR products and sequencing reaction conditions

gene region	primer	sequence (5' to 3')	PCR conditions				Reference
			Denaturation	Annealing	Extension	cycles	
barcode (1st half <i>Col</i>)	LCO, F	GGTCAACAAATCATAAAGATATTGG					N. Wahlberg
	Lep 3.1, R	AAATTTTAATTCCTGTTGGTACAGC	94°C for 30s	47°C for 30s	72°C for 1.5min	35	Dasmahapatra et al., in review
	Nancy, R	CCTGGTAAAAATAAAATATAAACTTC					N. Wahlberg
	K698=Rush, F	TACAATTTATCGCCTAAACTTCAGCC					Silva-Brandão et al. (2005), Brower et al. (2006)
	K699, R	WGGGGGGTAAACTGTTTCATCC					N. Wahlberg
	Ron, F	GGATCACCTGATATAGCATTCCC	95 for 1 min	45 for 1 min	72 for 1.5 min.	33	Brower et al. (2006)
<i>Col</i> (2nd half)	Jane, R	TAAAATTACTCCTGTTAATCCTCC					Brower et al. (2006)
	Wyman, R	GYTGAGCTCAWACAATAAATCCTA					Brower et al. (2006)
	Jerry, F	CAACAYTTATTTTGATTTTIGG	94°C for 30s	47°C for 30s	72°C for 1.5min	35	Simon et al. (1994), Mallarino et al. (2005)
	Pat, R	ATCCATTACATATAATCTGCCATA					Simon et al. (1994), Mallarino et al. (2005)
	Rudy, F	GAAGTTTATATTTTAATTTTACCGGG	95 for 1 min	45 for 1 min	72 for 1.5 min.	33	Brower and Jeansonne (2004)
<i>ColII</i>	Geoith, F	TAGGWTTAGCWGAATAACC	94°C for 45s	55°C for 45s	72°C for 1.5min	35	Dasmahapatra, unpub.
	Evaith, R	GAGACCAATACTTGCTTTCAGACATCT					
	GeorgeIII, F	TAGGTITAGCIGGAATACCTCG	95 for 1 min	45 for 1 min	72 for 1.5 min.	33	Brower and Jeansonne (2004)
	Imelda, R	CATTAGAAGTAATTGCTAATTTACTA					Brower et al. (2006)
	Eva, R	GAGACCAATACTTGCTTTCAGTCATCT					Brower and Jeansonne (2004)
	Strom, F internal	TAATTGAACTATYTTACCIG					Brower and Jeansonne (2004)
<i>EF1a</i>	Ef-M44-1, F	CTGAGCGYGARCGTGGTAT	94°C for 1min	55°C for 1min	72°C for 1.5min	35	Cho et al. (1995), Mallarino et al. (2005)
	EFrcM4, R	ACAGCVACKGTYTGYCTCATRTC					Cho et al. (1995), Mallarino et al. (2005)
	Ef1a-257F, F internal	TATCACTATTGACATCGC					
	40 (M3 of Cho in part), F	GTCGTSATYGGWCACGTMGATT	94 for 1 min	62 for 1 min.	72 for 1.5 min.	37	Cho et al. (1995)
	Gennifer, R	CGCACGGCAAAACGACCGAGRGG					Brower et al. (2006)
	52.6 (reM52.6 of Cho), R	GCYTCGTGGTGCATYTCAC					Cho et al. (1995)
	AL (M46-1 of Cho), F	GAGGAAATYAARAAGGAAG					Cho et al. (1995)

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References

- Brower, A. V. Z., Freitas, A. V. L., Lee, M. M., Silva-Brandao, K. L., Whinnett, A. & Willmott, K. R. 2006 Phylogenetic relationships among the Ithomiini (Lepidoptera: Nymphalidae) inferred from one mitochondrial and two nuclear gene regions. *Systematic Entomology* **31**, 288-301.
- Brower, A. V. Z., & Jeansonne M. M. 2004 Geographical populations and ‘subspecies’ of New World monarch butterflies (Nymphalidae) share a recent origin and are not phylogenetically distinct. *Ann. Entomol. Soc. Am.* **97**, 519-523.
- Cho, S., A. Mitchell, J. C. Regier, C. Mitter, R. W. Poole, T. P. Friedlander & Zhao S. 1995 A highly conserved nuclear gene for low-level phylogenetics: *elongation factor 1-alpha* recovers morphology-based tree for heliothine moths. *Mol. Biol. Evol.* **12**: 650-656.
- Dasmahapatra, K. K., Vásques, A. S., Chung, J. & Mallet, J. Genetic analysis of a wild-caught hybrid between non-sister *Heliconius* butterfly species. *Biology Letters*, *accepted*
- Mallarino, R., Bermingham, E., Willmott, K. R., Whinnett, A. & Jiggins, C. D. 2005 Molecular systematics of the butterfly genus *Ithomia* (Lepidoptera: Ithomiinae): a composite phylogenetic hypothesis based on seven genes. *Molecular Phylogenetics and Evolution* **34**, 625-644.
- Silva-Brandão, K. L., Freitas, A. V. L., Brower, A. V. Z. & Solferini, V. N. 1995. Phylogenetic relationships of the New World Troidini swallowtails (Lepidoptera: Papilionidae) based on COI, COII, and EF-1 α genes. *Molecular Phylogenetics and Evolution*, **36**, 468-483.
- Simon, C., Frati, F., Beckenbach, A., Crespi, B., Liu, H. & Flook, P., 1994. Evolution, weighting, and phylogenetic utility of mitochondrial gene sequences and a compilation of conserved polymerase chain reaction primers. *Annals of the Entomological Society of America* **87**, 651–702.
- Wahlberg, N. (<http://nymphalidae.utu.fi/Nymphalidae/Molecular.htm>)

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PCR were performed in 20 µl volume, with 2 µl template DNA, 0.4 U Bioline Taq Polymerase, 2 µl 10X buffer, 2 mM MgCl₂, 0.2 µM primer F, 0.2 µM primer R, 0.25 mM dNTP and ddH₂O.

PCR products were purified by adding 1U of shrimp alkaline phosphatase (SAP) and 1U of exonuclease I diluted in 1.5µl of USB SAP dilution buffer. Samples were incubated at 37°C for 40min, and the enzymes were inactivated at 80°C for 15min. Sequencing was carried out in both directions. One microliter of the purified PCR product was used as a template in a 10µl cycle sequence reaction containing 1µl Big Dye 3.1 (Applied Biosystems), 2µl 5X sequencing buffer (Applied Biosystems), 0.32 µM primer and ddH₂O.